## *Journal of Experimental Agriculture International*



*22(1): 1-7, 2018; Article no.JEAI.40439 ISSN: 2457-0591 (Past name: American Journal of Experimental Agriculture, Past ISSN: 2231-0606)*

# **Effect of Nitric Oxide on Seed Germination and Seedlings Development of Carrot under Water Deficit**

Guilherme Fontes Valory Gama<sup>1\*</sup>, Priscila Torres Cunha<sup>1</sup> **, Camila Andrade Fialho1 , Daniel Teixeira Pinheiro1\* and Igor Gonçalves de Paula1**

*1 Universidade Federal de Viçosa, Av. PH. Rolfs s/n, 36570000, Viçosa, MG, Brazil.*

## *Authors' contributions*

*This work was carried out in collaboration between all authors. Author GFVG designed the study, performed the statistical analysis, wrote the protocol, and wrote the first draft of the manuscript. Authors PTC and CAF managed the analyses of the study. Author DTP managed the literature searches. Author IGP worked translating Portuguese to English. All authors read and approved the final manuscript.*

## *Article Information*

DOI: 10.9734/JEAI/2018/40439 *Editor(s):* (1) Peter A. Roussos, Assistant Professor, Laboratory Pomology, Agricultural University of Athens, Greece. *Reviewers:* (1) Tauane Santos Brito, State University of Western Paraná, Brazil. (2) Manoj Kumar Yadav, SVP University of Agriculture & Technology Meerut, India. (3) Jackson da Silva, São Paulo State University, Brazil. Complete Peer review History: http://www.sciencedomain.org/review-history/23995

*Short Communication*

*Received 24th January 2018 Accepted 27th March 2018 Published 5th April 2018*

## **ABSTRACT**

Seed germination is strongly influenced by water deficit. Reduction of the osmotic potential and, consequently, of the water availability decreases the percentage of normal seedlings. It has been reported that nitric oxide (NO) is efficient in stimulating germination under both normal and stress conditions. The present work aims to evaluate the effect of exogenous application of NO, through a donor (SNP), on seed germination and on the development of carrot seedlings under water stress. Seeds of carrot cv. "Brasília" were submitted to water stress by PEG 6000, -0.3 MPa, and to SNP applications in the following concentrations: 100, 200 and 300 μM. The germinative assay was conducted at 20°C in germinator for 14 days. The first germination count (FGC), total germination (G), germination speed index (GSI), and dry matter of normal seedlings were evaluated. The SNP positively affected the germination and development of carrot seedlings under water stress. The most efficient concentration was observed at 100 μM.

\_

*\*Corresponding author: E-mail: pinheiroagroufv@gmail.com;*

*Keywords: Daucus carota L.; hydric stress; recovery; sodium nitroprusside.*

## **1. INTRODUCTION**

*.*

Abiotic stresses are classified as tensions caused by environmental conditions such as light, drought, radiation, heavy metals, hypoxia, temperature, and salinity [1]. It is estimated that abiotic stresses currently affect plant development as well as it decreases world agricultural production by up to 70% [2].

Germination is described as a three-phase process, initiated by seed imbibition (phase 1), activation of metabolic processes (phase 2), and root protrusion (phase 3) [3]. Thus, water is the most important factor regarding the germinative process, being responsible for rehydration of the tissues, intensification of respiration, and several metabolic processes that provide energy to the embryo [4].

Water availability decreases as the osmotic potential of the medium reduce, which causes negative impacts to both percentage and speed of germination [5]. Some osmotic agents exhibit similar behavior to that exerted by colloidal soil particles and can be used to simulate effects of water deficit on seeds [6]. For being chemically inert and non-toxic, polyethylene glycol (PEG 6000) is currently the most used solute to simulate water deficit once it keeps the seeds undamaged [7].

Aiming to recover the germination of seeds under water stress, the application of nitric oxide (NO) has been highlighted. NO is a gas considered as an outstanding mediator of intracellular and extracellular activities, regulating different physiological processes which attenuate effects of stresses, such as germination, antioxidative enzymes activity, breakage of dormancy, maturation, senescence, flowering, gravitropism, among others [8,9,10,11,12].

Sodium nitroprusside (SNP) is the most often NO donor used in germination studies due to its capacity to release this molecule when it is induced by the presence of light [13].

Due to the fact that it is usual the seedlings root system bifurcation in the soil, which makes them unfeasible, carrot plant (*Daucus carota* L.) is produced through direct seeding [14]. Moreover, it is considered as a high sensitive crop regarding drought conditions [15] and, therefore, studies related to water deficit at seed phase and

also the mechanisms involved in this process are of great importance.

Few reports are found regarding the action of NO on the recovery of carrot germination under such conditions. In this sense, this study's objective was to evaluate the effect of exogenous application of nitric oxide through sodium nitroprusside on seed germination and on the development of carrot seedlings under water deficit condition.

## **2. MATERIALS AND METHODS**

## **2.1 Location and Germination under Water Stress and Dry Matter of Seedlings**

The experiment was carried out at the Laboratory of Seeds Research, Plant Science Department, Universidade Federal de Viçosa, Brazil. Seeds of carrot (cultivar ''Brasilia''), produced by ISLA Ltda, was used in all tests.

Initially, preliminary germination tests were conducted on different PEG 6000 osmotic potentials, which were calculated according to the methodology of Villela et al. [7]. Based on such results, the potential of -0.3 MPa was selected in order to induce water stress in carrot seeds.

Seeds were placed on paper towel adhered to plastic "gerbox" boxes, moistened with PEG 6000 solutions at the osmotic potential of -0.3 MPa. The control was only treated with distilled water (0.0 MPa). The volume of solution was set as it corresponded to 2.5 times the dry paper weight. The gerbox boxes were kept in a germinator at 20°C. Results were expressed in percentage of normal seedlings obtained at the 14th day after sowing, as determined in the Rules for Seed Analysis [16]. First germination count (FGC) was performed alongside with the germination test, which consisted of the percentage of normal seedlings obtained at the 7th day after sowing [16].

The germination speed index (GSI) was performed through daily counts of normal seedlings, according to Maguire [17] (Equation 1).

Equation 1. Germination speed index (GSI)

$$
GSI = \left(\frac{G1}{N1} + \frac{G2}{N2} + \dots + \frac{Gn}{Nn}\right)
$$

Where:

GSI = germination speed index; G = number of normal seedlings counted;  $N =$  number of days from sowing to stabilization.

In regard to the evaluation of dry matter of seedlings, normal seedlings obtained from the germination tests were kept in paper bags and placed in a forced air-circulating oven at 75ºC for 48 hours. Subsequently, seedlings were weighed in a precision scale and the mean results expressed in grams.seedling<sup>-1</sup>.

#### **2.2 Effect of NO on Germination**

Besides the germination evaluated at 0 and -0.3 MPa, 3 treatments under stress (-0.3 MPa) were conducted containing SNP at concentrations of 100, 200 and 300 μM. Germination, GSI, and dry matter of seedlings were evaluated as abovementioned.

## **2.3 Experimental Design and Statistical Analysis**

The experiment was conducted in a completely randomized design with 5 treatments (control; stress; stress + 100 μM SNP; stress + 200 μM SNP; stress + 300 μM SNP) and four replications. Each experimental unit was composed of 50 seeds. Data were submitted to ANOVA and, subsequently, means were compared by the F test at 1% probability. Means of the different doses of SNP were adjusted to regression equations as well as their coefficients evaluated by the t-test at 1% and 5% probability in the SAS software.

#### **3. RESULTS AND DISCUSSION**

The analysis of variance showed significant effect for all variables. Thus, it is possible to infer that the water stress simulated by PEG 6000 solution caused negative effects on the germination of carrot seeds as well as on the development of seedlings. On the other hand, nitric oxide through SNP was able to partially recover both of them. By comparing results obtained by the control with the treatment submitted to water deficit, it is possible to state certain sensitivity of the carrot to water stress. Results of stress-only treatment were lower for all evaluations. As for the first count test, germination under the potential of -0.3 MPa was zero. According to Barroso [18], in unfavorable conditions, the germination time tends to increase until the seeds are able to develop adaptation mechanisms or until it is totally inhibited.

Carrot seeds regarding the control (0 MPa) and under water deficit (-0.3 MPa) presented germination rates of 82 and 22%, respectively. Such results show the drastic difference between the two treatments in regard to the percentage of normal seedlings (Fig. 1A).





*Means followed by different letters differ statistically by F test at 1% probability (P≤0.01)*

The predicted sensitivity to water stress condition is confirmed by Rodrigues et al. [19], who verified a reduction of carrot seed germination rates, also at the osmotic potential of -0.4 MPa, induced by PEG 6000 solution.

By the decrease of GSI, it is safe to say that the seeds germination speed was also affected by the osmotic potential of -0.3 MPa (Fig. 1B). Similarly, Silva et al. [20] observed a reduction in carrot seeds vigor at the potential of -0.3 MPa, which negatively impacted the GSI and germination speed.

Besides affecting imbibition, germination rates, and germination speed, water deficit also interferes with the seedlings growth. The stress condition can alter root meristem cells as well as the plant shoot system, interrupting division and cellular expansion along with cells turgescence. Such effects also harm seedlings development [21]. This fact explains the reduced dry matter

*Gama et al.; JEAI, 22(1): 1-7, 2018; Article no.JEAI.40439*

observed in the stress-treated plants as compared to control (Fig. 2).



#### **Fig. 2. Dry matter of carrot seedlings under water stress** *Means followed by different letters differ statistically by*

*F test at 1% probability (P≤0.01).*



**Fig. 3. First germination count (A), germination (B), and germination speed index (GSI) (C) of carrot seeds under water stress treated with SNP** *\*\*Equations are significant by F test at 1% probability (P≤0.01)*

Application of NO through the SNP donor increased first count and germination rates in relation to seeds submitted only to water stress (Fig. 3A and 3B). Several authors stated positive effects of NO on seed germination performance [22,23]. The causes may be related to the role of NO in the mobilization of β-amylase enzyme in seeds during germination [24].

In general, different concentrations of SNP slightly affected first germination count (Fig. 3A). Differently, as for germination rates, the concentration of 100 μM of SNP was the most efficient (Fig. 3B). Li et al. [25] found different results, when evaluating the effect of nitric oxide on parsley seeds under salt stress, pointed out an increase in germination rates at the concentration of 200 μM.

Seeds treated with SNP showed higher GSI values than those submitted only to stress (Fig. 3C). This fact supports the hypothesis that NO is efficient on recovering seed germination. However, there was no significant difference among the concentrations. Sarath et al. [23] and Liu et al. [26] state that the application of NO donors stimulates the seeds as they increase germination rates and also germination speed. It can be explained by the increase of α-amylase [27] and β-amylase activities [24], which promotes a greater and faster conversion of starch to soluble sugars [28].

Dry matter was positively influenced by the presence of SNP. The highest values were observed at the 100, 200 and 300 μM concentrations, respectively (Fig. 4).



**Fig. 4. Dry matter of carrot seedlings under water stress at different SNP concentrations**  *\*\*Equation is significant by F test at 1% probability (P≤0.01)*

As stated by Sarath et al. [23], NO targets proteins that act on elongation (radicle emergence) and cell growth (extension of coleoptiles) during the germination process. In a complementary way, Correa-Aragunde et al. [29] explain that SNP at low concentrations stimulates root growth, while high concentrations promote an inhibitory effect.

In general, the application of NO favored both germination and dry matter for carrot as compared to the stress treatment at -0.3 MPa. Such results corroborate with Pereira et al. [30], while in a study performed with *Plathymenia reticulata*, observed in pre-soaked seeds with SNP (100 μM) increases up to 155% in GSI. Furthermore, Kopyra & Gwóźdź [31] verified that *Lupinus luteus* seeds treated with NO increased germination rates by 20-40% after 18 and 24 hours.

## **4. CONCLUSION**

Application of SNP showed to be able of reduce physiological damage of carrot seeds under water stress. The highest values of germination and dry matter of seedlings were observed at the concentration of 100 μM.

#### **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

#### **REFERENCES**

- 1. Hirayama T, Shinozaki K. Research on plant abiotic stress responses in the post‐genome era: Past, present and future. The Plant Journal. 2010;61(6): 1041-1052. English.
- 2. Mantri N, Patade V, Penna S, Ford R, Pang E. Abiotic stress responses in plants: Present and future. In: Ahmad P, Prasad MNV, editors. Abiotic stress responses in plants: Metabolism, productivity and sustainability. Springer, New York; 2012. English.
- 3. Nonogaki H, Bassel GW, Bewley JD. Germination still a mystery. Plant Science. 2010;179(6):574-581. English.
- 4. Carvalho NM, Nakagawa J. Seeds: Science, technology and production. 5th ed. FUNEP: Jaboticabal; 2012. Portuguese.

*Gama et al.; JEAI, 22(1): 1-7, 2018; Article no.JEAI.40439*

- 5. Lopes JC, Pereira MD. Germination and development of jatropha seedlings under water stress conditions. Ciências Agrárias. 2011;32(1):1837-1842. Portuguese.
- 6. Parmar MT, Moore RP. Carbowax 6000, mannitol, and sodium chloride for simulating drought conditions in germination studies of corn *(Zea mays* L.) of strong and weak vigor. Agronomy Journal. 1968;60(2):192-195. English.
- 7. Villela FA, Doni-Filho L, Sequeira EL.. Table of osmotic potential as a function of polyethilene glycol 6000 concentration and<br>temperature. Pesquisa Agropecuária Pesquisa Agropecuária Brasileira. 1991;26(11/12):1957-1968. Portuguese.
- 8. Besson-Bard A, Pugin A, Wendehenne D. New insights into nitric oxide signallig in plants. Annual Review of Plant Biology. 2008;59:21-39. English.
- 9. Qiao W, Fan LF. Nitric oxide signaling in plant responses to abiotic stresses. Journal of Integrative Plant Biology. 2008;50(10):1238-1246. English.
- 10. Siddiqui MH, Al-Whaibi MH, Basalah MO. Role of nitric oxide in tolerance of plants to abiotic stress. Protoplasma. 2011;248(3): 447-455. English.
- 11. Fan HF, Du CX, Guo SR. Nitric oxide enhances salt tolerance in cucumber seedlings by regulating free polyamine content. Environmental and Experimental Botany. 2013;86:52-59. English.
- 12. Wang P, Du Y, Hou YJ, Zhao Y, Hsu CC, Yuan F, et al. Nitric oxide negatively regulates abscisic acid signaling in guard cells by S-nitrosylation of OST1. Proceedings of the National Academy of Sciences. 2015;112(2):613- 618. English.
- 13. Wang PG, Xian M, Tang X, Wu X, Wen Z, Cai T, et al. Nitric oxide donors: Chemical activities and biological applications. Chemical Reviews. 2002;102:1091-1134. English.
- 14. Souza, RJS, Assis, RP. Soil preparation and planting. In: Nick C; Borém, A, editors. Carrot from sowing to harvest. 1st ed. Viçosa: Editora UFV; 2016. Portuguese.
- 15. Marouelli WA, Oliveira RA, Silva WLC. Irrigation of carrot crops. Brasília: Embrapa hortaliças, circular técnica, 48; 2007. Portuguese.
- 16. Brasil. Ministério da agricultura e reforma agrária. Rules for seed analysis. Brasília: SNAD/DNDV/CLAV. 2009;395. Portuguese.
- 17. Maguire JD. Speed of germination-aid in selection and evaluation for seedling emergence and vigor. Crop Science. 1962;2(1):176-177. English.
- 18. Barroso CM, Franke LB, Barroso IB. Substratum and light on the germination of sinningia. Horticultura **Brasileira.** 2010;28:236-240. Portuguese.
- 19. Rodrigues DL, Lopes HM, Da Silva ER. Carrot seeds imbibition and germination under adverse environmental situations. Revista Científica Rural. 2017;19(2):205-216. Portuguese.
- 20. Silva MCDC, Medeiros AF, Dias DCDS, Alvarenga EM, Coelho FS, Braun H. Effect of heat and water stress on germination and vigour of carrot seeds. Idesia (Arica). 2011;29(3):39-44. Portuguese.
- 21. Hellal FA, El-Shabrawi HM, El-Hady MA, Khatab IA, El-Sayed SAA, Abdelly C. Influence of PEG induced drought stress on molecular and biochemical constituents and seedling growth of Egyptian barley cultivars. Journal of Genetic Engineering and Biotechnology. 2017;15(2). (in press). English.
- 22. Bethke PC, Gubler F, Jacobsen JV, Jones RL. Dormancy of arabidopsis seeds and barley grains can be broken by nitric oxide. Planta. 2004;219(5):847-855. English.
- 23. Sarath G, Bethke PC, Jones R. Nitric oxide accelerates seed germination in warmseason grasses. Planta. 2006;223(6): 1154-1164. English.
- 24. Zhang H, Shen WB, Zhang W. A rapid response of b-amylase to nitric oxide but not gibberellin in wheat seeds during the early stage of germination. Planta. 2005;220(5):708–716. English.
- 25. Li W, Liu X, Khan MA, Yamaguchi S. The effect of plant growth regulators, nitric oxide, nitrate, nitrite and light on the germination of dimorphic seeds of suaeda salsa under saline conditions. Journal of plant research. 2005;118(3):207–214. English.
- 26. Liu HY, Yu X, Cui DY, Sun MH, Sun WN, Tang ZC et al. The role of water channel proteins and nitric oxide signaling in rice seed germination. Cell Research. 2007;17(7):638–649. English.
- 27. Beligni MV, Fath A, Bethke PC, Lamattina L, Jones RL. Nitric oxide acts as an antioxidant and delays programmed cell death in barley aleurone layers. Plant Physiology. 2002;129(4):1642-1650. English.

*Gama et al.; JEAI, 22(1): 1-7, 2018; Article no.JEAI.40439*

- 28. Zhang H, Shen WB, Xu LL. Effects of nitric oxide on the germination of wheat seeds and its reactive oxygen species metabolisms under osmotic stress. Acta<br>Botanica Sinica. 2003;45(8):901-905. 2003;45(8):901-905. English.
- 29. Correa-Aragunde N, Graziano M, Lamattina L. Nitric oxide plays a central role in determining lateral root development in tomato. Planta. 2004;218(6):900-905. English.
- 30. Pereira BLC, De Lima e Borges EE, Oliveira AC, Leite HG, Gonçalves JDC.

Influence of nitric oxide on the germination of seeds of plathymenia reticulata benth<br>with low vigor Scientia Forestalis. with low vigor. Scientia 2010;38(88):629-636. English.

31. Kopyra M, Gwóźdź EA. Nitric oxide stimulates seed germination and counteracts the inhibitory effect of heavy metals and salinity on root growth of Lupinus luteus. Plant Physiology and<br>Biochemistry. 2003:41(11):1011-1017. Biochemistry. 2003;41(11):1011-1017. English.

 $\_$  , and the set of th *© 2018 Gama et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.*

> *Peer-review history: The peer review history for this paper can be accessed here: http://www.sciencedomain.org/review-history/23995*